

Effect of amino acid and peptide complex AB070597 on renal function in dogs with chronic kidney disease

The management of chronic kidney disease (CKD) in dogs remains a challenge to veterinary professionals and exerts financial burdens on their owners. Expanded knowledge and improvement in treatment would benefit all. The objective of the present study was to examine whether AB070597, a dietary-supplement compound of six amino acids and one peptide, can slow, halt, or reverse the decline of renal function in dogs with CKD. Five dogs with CKD were enrolled in a longitudinal study over 29 weeks. Animals received bi-daily oral doses of AB070597 from the point of enrolment to the end of the study, and blood samples were taken approximately every four weeks (for a total of 10 time intervals) to measure CKD-relevant biochemical parameters.

Results

The typical decline of renal function reported in dogs with CKD receiving standard palliative care, as measured by significant increases in blood-serum creatinine concentration (SCr), blood urea nitrogen (BUN), blood-serum phosphate concentration (PHOS), and lowered urine specific gravity (USG), was not observed. Median SCr, BUN, PHOS, and USG were stabilised and there was no significant change during any time interval.

Conclusions

These findings suggest that AB070597 may be a useful tool in stabilising or preventing the progression of CKD in some canines. The formulation warrants further study.

Keywords: Chronic kidney disease, renal insufficiency, AB070597, dogs, dietary

Introduction

CKD is the most common kidney disease in dogs⁶⁵. Prevalence, however, varies widely from 0.05⁶⁶, 0.9, 0.5–1.5 to 3.74%⁶⁸, depending on the source population and inclusion criteria.

CKD is less prevalent in dogs than in cats and the age at onset varies due to a number of breed-related diseases which affect the canine kidney. The mean age of dogs diagnosed with CKD is seven years⁶⁹. Irrespective of disease cause, it is widely accepted that most dogs diagnosed with CKD will progress inevitably to end stage disease and that BUN, SCr, PHOS will increase and USG will decrease significantly^{3,4,5}.

Many pathways lead to renal disease in humans, dogs, and other mammals. Mammalian renal architectures are similar, and even though there are some metabolic differences, it is possible to draw from the body of scientific information concerning mammalian renal disease, and with appropriate caution, apply those findings in an attempt to improve available treatment options for dogs with renal dysfunction. The most interesting findings in this context are related to disease progression monitoring via SCr, and data that suggests the role of reactive oxygen species (ROS) in renal cell damage.

Certain lipoproteins induce the formation of reactive oxygen species (ROS) in glomeruli and in arteries. Antioxidants

may prevent the damaging effects of these lipoproteins^{24,32,34,36,39,40–43,47,55}. Progressive injury results directly and indirectly from angiotensin II receptors via mediators of angiotensin-II-induced renal injury through transforming growth factor- β , fibroblast growth factor- β , tumor necrosis factor- α and platelet-derived growth factor^{5,6,7,11,25,31,54}. In addition, angiotensin increases oxidative stress, which causes a vasoconstrictor effect by increasing the catabolism of nitric oxide (NO)^{20,26,35,36}. Aldosterone is also a major contributor to the progression of CKD²⁶. All of these compounds promote the progression of CKD by enhancing cell growth, fibrosis and inflammation, which destroy tubulointerstitial tissues and glomeruli⁴.

Multiple approaches have been used to prevent the progression of renal disease, such as protein-restricted diets, the control of hypertension with angiotensin converting enzyme inhibitors, diet substitution of saturated fats with polyunsaturated fats, immunosuppressants such as mycophenolate mofetil, corticosteroids such as prednisone^{1,36,52,57,58} and morphogenic agents such as bone morphogenic protein-7 (BMP-7)⁶⁴. None of these treatments hold promise for halting or reversing disease progression, with the exception of BMP-7. Previous studies have shown that BMP-7 can reverse epithelial to mesenchymal transition in murine models of acute renal failure and can promote renal tissue repair⁶⁴. AB070597 is a cytoprotective agent that reduces damage to renal tubules and increases the glomerular filtration rate (GFR), stimulates gluconeogenesis and suppresses proteolysis in skeletal muscle, has strong anti-inflammatory properties, is a precursor for NO production and increases BMP-7 (Archer J, unpublished observations).

Methods and Subjects

An open-enrolment, open-ended study format was selected so that dogs with confirmed CKD could be added periodically. Dogs were confirmed with CKD if 1) their SCr \geq 2.3 mg/dL, 2) their USG was less than 1.050, 3) their BUN was elevated near or above the high end of the normal range, 4) their PHOS was elevated near or above the high end of the normal range and 5) their clinical history included signs attributable to CKD (i.e. persistent azotemia, chronic polyuria and polydipsia, or small kidneys on abdominal palpitation). If one or more of these signs were present with increased SCr, dogs were considered for inclusion in the study. Dogs with suspected or verified conditions, such as pyelonephritis, uncontrolled hypothyroidism, acute renal failure, cancer, or other CKD-unrelated diseases were excluded. The criteria used to establish the cessation of progressive renal injury were 1) a halt in the rise of SCr, BUN and PHOS and a halt to the decline of USG, all for an extended period of time. No statistically significant deterioration in these values would signify no statistically significant disease progression. Dog owners were given informed consent forms for review and acceptance. The amino acids and peptide in AB070597 were purchased from Spectrum Chemical Company (Gardena, California).

Over a 29-week period, five subjects ranging in weight from 3.2 to 18.5 Kg (mean = 7.8, median = 4.7, range = 3.2–18.5 Kg), all on non-protein-restricted commercial diets, received two 1000 mg / 4.5 Kg body weight oral daily doses of AB070597 as a dietary supplement. Doses were mixed with approximately 3

millilitres of water and administered directly into each subject's mouth, or the dose was sprinkled directly on a small amount of food and fed to the subject. AB070597 was readily accepted without rejection. SCr, BUN, PHOS and USG measurements were made for each subject at four-week intervals during the 29-week study (mean = 22, median = 16, range = 16-38 weeks).

Statistical Analysis

Statistical comparisons were made by comparing measured parameters upon entering the study with measured parameters at the end of the study. Values were calculated with the Wilcoxon signed-rank test (VassarStats: Website for Statistical Computation, www.vassarstats.net). Differences were considered statistically significant at $P \leq 0.05$ (two tailed).

Results

Each patient was monitored over the course of the study. Assessments were made for general body condition, weight change and ease of administration of AB070597. There were no owner complaints or concerns regarding administration of the supplement. General body condition, including coat, appearance and grooming habits, improved in each patient, and 60% gained weight, while body weight of the other 40% remained stable without further loss. There were no reports of gastrointestinal upset or diarrhoea. The medians of SCr, BUN, PHOS and urine USG showed no statistically significant change from start to finish Table 1.

Discussion

Measured parameter	Start median	Start IQR	End median	End IQR	P
SCr (mg/dl)	2.0	1.4	1.6	1.9	ns
BUN (mg/dl)	48	54	51	44	ns
PHOS (mg/dl)	3.4	1.6	1.5	2.3	ns
USG	1.015	0.004	1.022	0.006	ns

IQR = Interquartile range

Table 1. Changes in the measured parameters of the sample population over the study duration.

CKD is a progressive disease that results in significantly increased SCr, BUN, PHOS and lowered USG over time. Oral administration of AB070597 attenuated progression, such that there were no significant (ns) changes in those values. A possible hypothesis is that these results are due to biochemical effects of the individual components of AB070597.

L-arginine protects renal tissue from the negative effects of renal ischemia^{12, 50, 53, 59} and facilitates the disposal of protein and metabolic waste, and acts globally on muscle metabolism, vascular tone regulation and immune system function, and promotes the release of numerous hormones (glucocorticoids, growth hormone, prolactin, insulin, somatostatins, glucagons, catecholamines) through various pathways, whose disturbance can cause detrimental effects on renal function^{2, 3, 4, 18}. L-arginine administration also relieves a variety of pathological states, including kidney hypertrophy and glomerular thrombosis^{2, 10, 50, 53}.

Glycine is produced in the mammalian body in amounts up to fifty times greater than those taken in daily by diet. Any decrease in its natural production is therefore of concern. The body uses this amino acid to form RNA, DNA porphyrin, bone collagen, glutathione, heme, bile, and salts and for the detoxification and conjugation of toxic products, both

exogenous and endogenous. It exerts a cytoprotective effect against anoxia, ischemia, heat, antibiotics, metals, and indomethacin-induced kidney damage. It also increases the GFR and thereby improves kidney function^{16, 17, 48}.

L-glutamine is the precursor of nucleotides and proteins and is the substrate for and stimulates gluconeogenesis in all organs and tissues. This amino acid regulates carbohydrate metabolism and suppresses proteolysis in skeletal muscles and stimulates protein synthesis, thereby counteracting the muscle-wasting effects caused by CKD^{8, 21, 23, 37, 38, 46, 56, 61, 62}. L-glutamine is also an efficient ROS scavenger⁶³.

L-histidine concentrates in the brain at a level five times higher than in blood serum. It is readily available from food, but food intake is reduced in animals with CKD, thereby reducing the normal total cerebral amount. It also has anti-inflammatory properties and counteracts the damaging effects of ROS formed by those processes^{15, 44}.

L-aspartic acid and L-glutamic acid, as sodium salts, are present in all tissues. The highest concentrations are located in the central nervous system. L-aspartic acid sodium is distributed throughout the central nervous system and spinal cord²². L-glutamic acid sodium is distributed in the caudate nucleus and cerebral cortex³⁰. They act as efficient ROS scavengers and thereby protect the kidneys from ongoing damage caused by these ROS radicals⁶³.

L-carnosine increases the production of BMP-7²⁸. BMP-7 induces mesenchymal-to-epithelial transition in renal fibroblasts and facilitates regeneration of injured kidney⁶⁴.

Conclusion

Dogs with CKD treated with AB070597, as a dietary supplement, did not experience increased SCr, BUN, PHOS, or a decrease in USG typically seen in dogs with CKD receiving standard palliative care. Treated animals had stable or reduced values for these parameters for up to 29 weeks. Results suggest that the oral supplement AB070597 may promote the maintenance of renal function and potentially attenuate or prevent progression CKD in dogs. Additional studies with more subjects and longer follow-ups are warranted, and future studies of AB070597 and its effect on renal function in dogs employing a much larger population are planned.



REFERENCES

1. Barsotti G, Cupisti A, Gervasi GB et al. Effects of oral administration of heparan sulfate in the rat remnant kidney model. *Nephron* 1999; 81: 310–318.
2. Bode-Boger SM, Boger RH, Kienk S et al. Chronic dietary supplementation with L-arginine inhibits platelet aggregation and thromboxane-A synthesis in hypercholesterolemic rabbits in vivo. *Cardiovasc Res* 1998; 17: 756–764.
3. Boger RH, Bode-Boger SM, Kienke et al. Dietary L-arginine decrease myointimal cell proliferation and vascular monocyte accumulation in cholesterol-fed rabbits. *Atherosclerosis* 1998; 136: 67–77.
4. Boger RH, Bode-Boger SM, Muggle A et al. Supplementation of hypercholesterolemic rabbits with L-arginine reduces the vascular release of superoxide anions and restores NO production. *Atherosclerosis* 1995; 117: 273–284.
5. Cambese VM. Neurogenic factors and hypertension in renal disease. *Kidney Int* 2000; 57(Suppl 75): S-2–S-6.
6. Campese VM, Kogosov E, Koss M. Renal afferent denervation prevents the progression of renal disease ablation model of chronic renal failure in the rat. *Am J Kidney Dis* 1995; 26: 861–865.
7. Campese VM, Romo MS, Levitan D, Massry SG. Mechanisms of autonomic nervous system dysfunction in uremia. *Kidney Int* 1981; 20: 246–253.
8. Cercosimo E, Williams P, Hoxworth B et al. Glutamine blocks lipolysis and ketogenesis of fasting. *Am J Physiol* 1989; 250: E248–E252.
9. Chunsen D, Junwei Y, Youhua L. Single injection of naked plasmid encoding hepatocyte growth factor prevents cell death and ameliorates acute renal failure in mice. *J Am Soc Nephrol*. 13: 411–422, 2002.
10. Clarkson P, Adams MR, Powe AJ et al. Oral L-arginine improves endothelium-dependent dilation in hypercholesterolemic young adult. *J Clin Invest* 1996; 97: 1989–1994.
11. Converse RI, Jacobsen TN, Toto RD et al. Sympathetic overactivity in patients with chronic renal failure. *N Engl J Med* 1992; 327: 1912–1918.
12. Cooke JP, Singer AH, Tsao PS et al. Antithrombotic effects of L-arginine in the hypercholesterolemic rabbit. *J Clin Invest* 1992; 90: 1168–1172.
13. Covic A, Caruntu ID, Marian D, Volovat C, Ghiciuc C, Costin C, Florea L, Cotutiu C, Covic M. Prognosis and treatment of membranous glomerulonephritis—A 5 year prospective study. *Rev Med Chir Soc Med Nat Lasi*. 2002 Apr-Jun; 104(2): 63–74.
14. D'Amico G. Tubulointerstitium as predictor of progression of glomerular disease. *Nephron* 1999; 83: 289–295.
15. Decker EA, Ivanov V, Zhu BJ, Frei B. Inhibition of low-density lipoprotein oxidation by carnosine. *J Agric Food Chem* 2001; 49: 511–516.
16. Doolan PD, Harper HA, Hutchin ME, Alpen EL. The renal tubular response to amino acid loading. *J Clin Invest* 1956; 35: 888–896.
17. Doolan PD, Harper HA, Hutchin ME, Shreeve WW. Renal clearance of eighteen individual amino acids in human subjects. *J Clin Invest* 1955; 34: 1247–1255.
18. Drexler H, Zeiher AM, Meinzer K, Just H. Connection of endothelial dysfunction in coronary microcirculation of hypercholesterolemic patients by L-arginine. *Lancet* 1991; 338: 1546–1550.
19. Eddy AA, Liu E, McCulloch L. Interstitial fibrosis in hypercholesterolemic rats: role of oxidation, matrix synthesis, and proteolytic cascades. *Kidney Int* 1998; 53: 1182–1189.
20. Fogo A. Glomerular hypertension, abnormal glomerular growth, and progression of renal diseases. *Kidney Int* 2000; 57(Suppl 75): S-15–S-21.
21. Garrison R. Lysine, Tryprophan and other amino acids. Copyright 1982 by Keats Publishing, Inc. Printed in United States of America, 27 Pine Street.
22. Graham LT, Shank RP, Werman R, Aprison MH. Distribution of some synaptic transmitter suspects in cat spinal cord glutamic acid, aspartic acid, c-amiobutyric acid, glycine, and glutamine. *J Neurochem* 1967; 14: 465–472.
23. Hankard RG, Darmaun D, Sager BK et al. Response of glutamine metabolism to exogenous glutamine in humans: *Am J Physiol* 1995; 269: E663–E670.
24. Hebert LA, Kusek JW, Greene T et al. Effects of blood pressure control on progressive renal disease in blacks and whites. Modification of Diet in Renal Disease Study Group. *Hypertension* 1997; 30: 428–435.
25. Herzog CA, Ma JZ, Collins AJ. Poor long-term survival after acute myocardial infarction among patients on long-term dialysis. *N Engl J Med* 1998; 339: 799–805.
26. Ibrahim HN, Rosenberg ME, Greene EL et al. Aldosterone is a major factor in the progression of renal disease. *Kidney Int* 1997; 52(Suppl 63): S-115–S-119.
27. Ito K, Chen J, Vaughan ED Jr. et al. Dietary L-arginine supplementation improves the glomerular filtration rate and renal blood flow after 24 hours of unilateral ureteral obstruction in rats. *J Urol*, 2004; Feb; 171(2 Pt 1): 926–30.
28. Ito-Kato E, Suzuki N, Maeno M, Takada T, Tanabe N, Takayama T, Ito K, Otsuka K. 2004. Effect of carnosine on runt-related transcription factor-2/core binding factor alpha-1 and Sox9 expressions of human periodontal ligament cells. *J Periodontol Res*. 39(3):199–204.
29. Izzo ZI, Izzo MS, Sterns RH, Freeman RB. Sympathetic nervous-system hyperactivity in maintenance hemodialysis patients. *Trans Am Soc Artif Organs* 1982; 28: 604–606.
30. Johnson JI, Aprison MH. The distribution of glutamate and free amino acids in thirteen specific regions of the rat central nervous system. *Brain Res* 1971; 26: 141–148.
31. Katholi RE, Whitlow PI, Hageman GR, Woods T. Intra-renal adenosine produces hypertension by activating the sympathetic nervous system via the renal nerves. *J Hypertension* 1984; 2: 349–359.
32. Keane WF: The role of lipid in renal disease: future challenges. *Kidney Int* 2000; 57(Suppl 75): S-27–S-31.
33. Kehrer G, Blech M, Kallerhoff M, Langheinrich M et al. Contribution of amino acids in protective solutions to postischemic functional recovery of canine kidneys. *Res Exp Med (Berl)*; 1989; 189(6): 381–96.
34. Klahr S, Levey AS, Berk GJ et al. The modification of diet in renal disease (MDRD) Study Group. The effects of dietary protein restriction and blood pressure control on the progression of chronic renal disease. *N Engl J Med* 1994; 330: 877–884.
35. Klahr S, Morrissey JJ. The role of vasoactive compounds, growth factors and cytokines in the progression of renal disease. *Kidney Int* 2000; 57(Suppl 75): S-7–S-14.
36. Klahr S. Prevention of progression of nephropathy. *Nephrol Dial Transplant* 1997; 12(Suppl 2): 63–66.
37. Kreider M, Stumvoll M, Mever C et al. Steady state and non steady measurement of plasma glutamine turnover in humans. *Am J Physiol* 1997; 272: E621–E627.
38. Lavoinne A, Baquet A, Hue L. Stimulation of glucogen synthesis and lipogenesis by glutamine in isolated rat hepatocytes. *Biochem J* 1987; 248: 429–437.
39. Lazarus JM, Bourcoignie JJ, Buckalew VM et al. For the Modification of Diet in Renal Disease Study Group. Achievement of safety of a low blood pressure goal in chronic renal disease in the Modification of Diet in Renal Disease Study. *Hypertension* 1997; 29: 641–650.595.
40. Levey A, Adler S, Caggiula A et al. For The MDRD Study Group. Effects of dietary protein restriction on the progression of advanced renal disease in the Modification of Diet in Renal Disease (MDRD) Study. *Am J Kidney Dis* 1996; 27: 652–663.
41. Levey AS, Greene T, Schluchter MD et al. The modification

- of diet in renal disease (MDRD) study and the diabetes control and complications trial (DCCT) research group: glomerular filtration rate measurement in clinical trials. *J Am Soc Nephrol* 1993; 3: 1159–1171.
42. Beck GJ, Beck RL, Coggins CH, Gassman JJ, Hunsicker LG, Schluter MD, Williams GW. Modification of Diet in Renal Disease (MDRD) study group. Design and statistical issues of the Modification of Diet in Renal Disease Trial. *Controlled Clin Trials* 1991; 12: 566–586.
 43. Greene T, Bourgoignie JJ, Habwe VQ, Kusec JW, Snetsellaar L, Soucie JM, Yamamoto M. Modification of Diet in Renal Disease (MDRD) study group. Baseline characteristics in the Modification of Diet in Renal Disease Study. *J Am Soc Nephrol* 1993; 3: 1819–1834 (original version); 1993; 4: 1221–1236 (corrected version).
 44. Moncada S, Palmer RM, Higgs EA. Nitric oxide: Physiology, pathophysiology, and pharmacology. *Pharmacol Rev* 1991; 43: 109–142.
 45. Niaudel P, Habib R. Methylprednisolone pulse therapy in the treatment of severe forms of Schonlein-Henoch purpura nephritis. *Pediatr Nephrol*. 1998 Apr; 12(3): 238–43.
 46. Perriello G, Nurjhan N, Stumvoll M et al. Regulation of gluconeogenesis by glutamine in normal postabsorptive humans. *Am J Physiol* 1997; 272: E437–E445.
 47. Peterson J, Adler S, Burkart J et al. For The Modification of Diet in Renal Disease (MDRD) study group. Blood pressure control, proteinuria, and the progression of renal disease: the Modification of Diet in Renal Disease Study. *Ann Intern Med* 1995; 123: 754–762.
 48. Pitts R. The effect of infusing glycine and of varying the dietary protein on renal hemodynamics in dog. *Am J Physiol* 1944; 142: 355–365.
 49. Pozzi C, Del Vecchio L, Locatelli F. Immunosuppressive therapy in IgA glomerulonephritis with chronic renal failure; case study presentation and literature review. *G Ital Nefrol*. 2002 Sep–Oct, 19 (5): 528–8.
 50. Reyes AA, Karl IE, Klahr S. Role of arginine in health and in renal diseases. *Am J Physiol* 1994; 267: F331–F346.
 51. Romero F, Rodriguez-Iturbe B, Parra G et al. Mycophenolate mofetil prevents the progressive renal failure induced by 5/6 renal ablation in rats. *Kidney Int* 1999; 55: 945–955.
 52. Romero F, Rodriguez-Iturbe B, Parra G et al. Mycophenolate mofetil prevents the progressive renal failure induced by 5/6 renal ablation in rats. *Kidney Int* 1999; 55: 945–955.
 53. Rossitch E, Alexander E, Black PM, Cooke JP. L-arginine normalizes endothelium function in cerebral vessels from hypercholesterolemic rabbits. *J Clin Invest* 1991; 87: 1295–1299.
 54. Rump LC, Amann K, Orth S, Ritz E. Sympathetic over-activity in renal disease: a window to understand progression and cardiovascular complications of uremia? *Nephrol Dial Transplant* 2000; 15: 1735–1738.
 55. Schena FP. Management of patients with chronic kidney disease. *Intern Emerg Med*. 2011; Oct;6 Suppl 1:77–83.
 56. Stumvoll M, Meyer C, Kreider M et al. Effects of glucagon on renal and hepatic glutamine gluconeogenesis in normal postabsorptive humans. *Metabolism* 1998; 47: 1227–1232.
 57. Tarif N, Bakris GL. Preservation of renal function: the spectrum of effects by calcium-channel blockers. *Nephrol Dial Transplant* 1997; 12: 2244–2250.
 58. ter Wee PM, Donker AJ. Clinical strategies for arresting progression of renal disease. *Kidney Int* 1992; 42(Suppl 38): S-114–S-120.
 59. Tsao PS, McEvoy LM, Drexler H et al. Enhanced endothelial adhesiveness is attenuated by L-arginine. *Circulation* 1994; 89: 23176–2182.
 60. Tuttle K, Puhlman M, Cooney S, Short R. Effects of amino acids and glucagons on renal hemodynamics in type 1 diabetes. *Am J Physiol Renal Physiol* 2002; 282: F103–F112.
 61. Varnier M, Leese GP, Thompson J, Rennie MJ. Stimulatory effect of glutamine on glycogen accumulation in human skeletal muscle. *Am J Physiol* 1995; 269: E309–E315.
 62. Vu G, Thompson JR. The effect of glutamine on protein turnover in chick skeletal muscle in vitro. *Biochem J* 1990; 265: 593–598.
 63. Yatzidis H. A new, superior, single and stable, amino acid and bicarbonate-based, glucose-free solution for peritoneal dialysis. *Dialysis and Transplantation* 2002; 31: 143–150.
 64. Zeisberg M, Hanai J, Sugimoto H, Mammoto T, Charytan D, Strutz F, Kalluri R. BMP-7 counteracts TGF-beta1-induced epithelial-to-mesenchymal transition and reverses chronic renal injury. *Nat Med* 2003; 9 (7): 964–8.
 65. Polzin DJ, Osborne CA, Ross S. Chronic kidney disease. In: Ettinger SJ, Feldman EC, eds. *Textbook of Veterinary Internal Medicine: Diseases of the Dog and Cat*, 6th ed. Stephen J. Ettinger, Edward C. Feldman, eds. St Louis, MO; Oxford: Elsevier Saunders; 2005:1756–1785.
 66. Macdougall DF, Cook T, Steward AP et al. Canine chronic renal disease: prevalence and types of glomerulonephritis in the dog. *Kidney Int* 1986;29:1144–1151.
 67. Brown SA. Management of chronic kidney disease. In: Elliott J, Grauer GF, eds. *BSAVA Manual of Canine and Feline Nephrology and Urology*, 2nd ed. Quedgeley: British Small Animal Veterinary Association; 2007:223–230.
 68. Sosnar M. Retrospective study of renal failure in dogs and cats admitted to University of Veterinary and Pharmaceutical Sciences Brno during 1999–2001. *Acta Veterinaria Brno* 2003;72:593–598.
 69. Shaw HS, Ihle, SL. *Small Animal Internal Medicine*, Darrin Kiessling, ed., Lippincott Williams and Wilkins, Baltimore, Maryland, USA, 1997.



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